

## A Revised Structure for Crotoamosmin from *Crotolaria ramosissima*

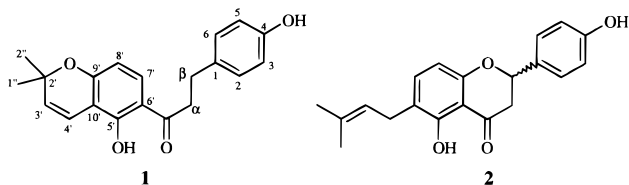
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The structure of crotoamosmin has been reassigned to 1-(5-hydroxy-2,2-dimethyl-2*H*-chromen-6-yl)-3-(4-hydroxyphenyl)-propanone (**1**) as determined by extensive NMR investigation.

Several years ago, the isolation and structure assignment of crotoamosmin from *Crotolaria ramosissima* Roxb. (Fabaceae) was reported.<sup>1</sup> Due to limited spectroscopic facilities available at that time, this compound was erroneously assigned to the flavanone structure **2** with unknown absolute configuration. We have reexamined this compound from the same plant source by modern NMR methods and found that it is rather a novel dihydrochalcone, namely, 1-(5-hydroxy-2,2-dimethyl-2*H*-chromen-6-yl)-3-(4-hydroxyphenyl)-propanone (**1**). Dihydrochalcones are rare natural products, and this is the first report of the isolation of such a compound from *Crotolaria* species.



The elemental formula of **1** was determined as C<sub>20</sub>H<sub>20</sub>O<sub>4</sub> by HRMS. The complete <sup>1</sup>H and <sup>13</sup>C connectivity was established by extensive use and interpretation of 2D (<sup>1</sup>H–<sup>1</sup>H) COSY, HMQC (one-bond <sup>13</sup>C–<sup>1</sup>H correlation), and HMBC (long-range <sup>13</sup>C–<sup>1</sup>H correlation) NMR spectra.<sup>2</sup> This provided unequivocally the atomic network for **1** (Table 1).

A few structural features are worth mentioning. The existence of a CH<sub>2</sub>–CH<sub>2</sub> group was obvious; <sup>1</sup>H NMR signals appeared as triplets, and the corresponding carbons were identified as CH<sub>2</sub> by a DEPT experiment. This group is flanked by a *p*-hydroxylated phenyl group at one end and by a carbonyl at the other. This was confirmed by the carbonyl chemical shift of δ<sub>C</sub> 206.1, a value that is much higher than in the carbonyl groups of chalcones but fits nicely in a dihydrochalcone.<sup>3</sup> The second phenyl group was found to be tetrasubstituted with two *ortho*-positioned hydrogens (H-7' and H-8'). The pyran ring contains a *cis*-configured double bond, as evidenced by the 10.0-Hz <sup>1</sup>H–<sup>1</sup>H coupling

**Table 1.** <sup>1</sup>H and <sup>13</sup>C NMR Chemical Shifts of **1**, Including <sup>13</sup>C–<sup>1</sup>H Long-Range Correlations<sup>a</sup>

position	<sup>1</sup> H	<sup>13</sup> C	HMBC C → H
1		133.2	
2	7.02	130.5	4, 6, β
3	6.69	116.4	1, 4, 5
4		156.8	
5	6.69	116.4	1, 3, 4
6	7.02	130.5	2, 4, β
α	3.11	41.0	β, 1, C=O
β	2.86	30.9	α, 1, 2/6, C=O
C=O		206.1	
2'		78.8	
3'	5.59 <sup>b</sup>	129.5	2', 10'
4'	6.63 <sup>b</sup>	116.7	2', 5'
5'		160.9	
6'		114.7	
7'	7.54	132.7	5', 9', C=O
8'	6.25	109.4	6', 10'
9'		160.6	
10'		110.3	
1''	1.39	28.7	2', 3', 2''
2''	1.39	28.7	2', 3', 1''
5'-OH <sup>c</sup>	13.1		

<sup>a</sup> HMBC, optimized to 7 Hz; solvent, CD<sub>3</sub>OD. <sup>b</sup> Vicinal <sup>1</sup>H–<sup>1</sup>H coupling constant *J* = 10.0 Hz. <sup>c</sup> Taken from CDCl<sub>3</sub> solution.

constant and a dimethyl carbinol group (δ<sub>C</sub> 78.8). It is attached to C-10' as proven by the observation of a long-range correlation between H-4' and C-5', whereas the carbonyl group is fixed at C-6' (H-7'/C=O correlation). The C-5' hydroxy group is involved in a hydrogen bridge as shown by the <sup>1</sup>H chemical shift (measured in CDCl<sub>3</sub>; δ<sub>C</sub> 13.1).

### Experimental Section

**General Experimental Procedures.** <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded in CDCl<sub>3</sub> using a Bruker DRX-500 spectrometer, and standard Bruker software for the 2D NMR experiments was used. The HRMS spectrum was obtained with a Varian MAT CH-5 mass spectrometer using electron impact.

**Plant Material.** As described previously.<sup>1</sup>

**Extraction and Isolation.** As described previously.<sup>1</sup>

**Compound 1:** <sup>1</sup>H and <sup>13</sup>C NMR data, see Table 1; EIMS *m/z* 324 (M<sup>+</sup>, 20), 309 (100), 291 (5), 203 (29), 187 (9), 185 (9), 161 (4), 160 (5), 107 (17), 91 (3), 77 (7). HRMS, *m/z* (M<sup>+</sup>), calcd for C<sub>20</sub>H<sub>20</sub>O<sub>4</sub>, 324.13615; found 324.13557.

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### References and Notes

- (1) Khalilullah, M.; Sharma, V. M.; Rao, P. S.; Raju, K. R. *J. Nat. Prod.* **1992**, *55*, 229–231.
- (2) Bruker AVANCE standard software.
- (3) Agrawal, P. K. *Carbon-13 NMR of Flavonoids*, Elsevier Science: Amsterdam, 1989.

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